

1 Meatal Swabs Contain Less Cellular Material and are Associated with a Decrease in
2 Gram Stain Smear Quality Compared to Urethral Swabs in Men

3

4 Stephen J. Jordan^{1#}, Jane R. Schwebke¹, Kristal J. Aaron¹, Barbara Van Der Pol¹,
5 Edward W. Hook III¹

6 ¹Department of Medicine, University of Alabama at Birmingham School of
7 Medicine, Birmingham, AL

8

9

10 **Running Title:** Meatal swabs perform poorly in Gram stain testing

11

12 #Address correspondence to Stephen J. Jordan: University of Alabama at
13 Birmingham, 1720 2nd Avenue South, THT 229, Birmingham, AL 35294, USA.

14 Phone: 1 (205) 934-5191. Fax: 1 (205) 934-5155. E-mail: sjordan@uabmc.edu

15

16 **Abstract:**

17 Urethral swabs are the test of choice for point-of-care Gram stain testing to diagnose
18 *Neisseria gonorrhoeae* (NG) and non-gonococcal urethritis (NGU) in men. As an
19 alternative to urethral swabs, meatal swabs have been recommended for collection of
20 urethral discharge to diagnose NG and *Chlamydia trachomatis* (CT) in certain
21 populations by nucleic acid amplification testing (NAAT), as they are a less invasive
22 collection method. However, as meatal swabs could be sampling a reduced surface
23 area and result in fewer collected epithelial cells when compared to urethral swabs, the

24 adequacy of meatal swab specimens to collect sufficient cellular material for Gram stain
25 testing remains unknown. We enrolled 66 men who received either a urethral swab or a
26 meatal swab and compared the cellular content and Gram stain failure rate. We
27 measured the difference in swab cellular content using the Cepheid Xpert® CT/NG
28 sample adequacy control crossing threshold and determined the failure rate of gram
29 stain smears (GSS) due to insufficient cellular material. Meatal smears were associated
30 with a significant reduction in cellular content ($P = 0.0118$), which corresponded with a
31 significantly higher GSS failure rate compared to urethral swabs (45% vs. 3%
32 respectively, $P < 0.0001$), in the absence of discharge. When discharge was present,
33 there was no difference between urethral and meatal swabs. Therefore, if GSS testing
34 is being considered for point-of-care diagnosis of NG or NGU in men, meatal swabs
35 should be avoided in the absence of a visible discharge.

36

37 **Introduction**

38 *Chlamydia trachomatis* (CT) and *Neisseria gonorrhoeae* (NG) infections continue to rise
39 with 1.4 million and > 350,000 cases reported, respectively, in the United States in 2015
40 (1); though non-gonococcal urethritis (NGU) remains the most common form of
41 urethritis in men (2). For screening at-risk individuals, the Centers for Disease Control
42 and Prevention (CDC) recommends highly sensitive nucleic acid amplification testing
43 (NAAT) of urine or a urethral swab of urethral secretions (3). Meatal swabs have been
44 suggested as a less invasive alternative to urethral swabs for specimen collection for
45 NAAT testing and are amenable to patient collection by eliciting less discomfort (4).
46 Although not yet FDA-approved or CDC-recommended for NAAT testing in men, meatal

47 swabs are recommended for NAAT testing in prepubertal boys with discharge, given
48 concerns about urethral trauma from urethral swabs (3,7). In men, self-obtained meatal
49 swabs appear to be equivalent to clinician-collected urethral swabs for diagnosing
50 CT/NG infection by NAAT, although there are mixed reports of the sensitivity of this
51 sample type (5, 6). Currently, no rapid (< 30 minutes) point-of-care NAAT test is
52 available to diagnose NG or NGU; and Gram stain smear (GSS) testing of urethral
53 secretions remains the test of choice in settings where rapid diagnosis is needed and
54 microscopy can be performed. The 2015 Sexually Transmitted Diseases Treatment
55 Guidelines specify that GSS testing of male urethral secretions is appropriate for
56 diagnosing NG or NGU, though no preferred swab type is indicated for specimen
57 collection. Meatal swabs are recommended as an alternative for collecting urethral
58 secretions in specific populations (7), though they have yet to be approved for NAAT
59 testing in men. In contrast to NAAT, which amplifies nucleic acids from lysed cells, GSS
60 testing requires collection of intact cells. As meatal swabs may sample a smaller
61 surface area than urethral swabs, and also sample a higher proportion of cornified
62 squamous epithelium from the meatus, it is possible that the number of intact cells
63 collected using a meatal swab may be insufficient to reliably diagnose NG or NGU by
64 GSS testing. To address this concern, we enrolled both symptomatic and asymptomatic
65 men and systematically assigned them to receive either a meatal or urethral swab and
66 measured the swab cellular content using the Cepheid Xpert® CT/NG sample
67 adequacy control crossing threshold (SAC_{CT}) (8) and also determined the failure rate of
68 GSS testing for each swab type. The SAC_{CT} is an internal control of the Xpert® CT/NG
69 assay and denotes the cycle number at which human hydroxymethylbilane synthase, a

70 single-copy housekeeping gene, is first detected by real-time PCR amplification.
71 Included in each assay to ensure specimen sample adequacy (9), the SAC_{CT} is
72 inversely proportional to the amount of cellular material present in the specimen. The
73 primary outcome of our study was to determine if meatal swabs were associated with a
74 lower cellular content and a higher GSS failure rate, compared with urethral swabs. A
75 secondary objective was to determine how the meatal swab was influenced by the
76 presence or absence of discharge.

77

78 **Results**

79 66 men were included in this study (Table 1). Participants were 20 to 69 (mean
80 29) years of age, and 56 (89%) were black. 27 men (41%) had visible discharge on
81 genital examination. Swab collection (meatal vs. urethral) was alternated such that 33
82 men provided a urethral swab and 33 men had a meatal swab taken. No difference in
83 age, race, symptoms, prior STI history, or number of sex partners in the last 30 days
84 was identified between the groups. 9 (14%) men had Gram-negative intracellular
85 diplococci (GNID) present on GSS and 20 men (30%) were diagnosed with NGU. The
86 Xpert® CT/NG assay, performed on both swab and voided urine specimens, diagnosed
87 20 men with CT and/or NG: 8 men were positive for CT alone, 4 men for both CT and
88 NG, and 8 men for NG alone.

89 To assess whether meatal swabs were associated with a decrease in cellular
90 content, we compared the SAC_{CT} of meatal swabs to urethral swabs. Meatal swabs
91 were associated with significantly higher SAC_{CT} values, indicating they contained less
92 cellular material, compared to urethral swabs (mean 25.6 vs 23.9, $P = 0.0026$, Table 1).

93 This difference in cellular content did not affect the performance of the NAAT since the
94 swab sample NAAT results were 100% concordant with the urine NAAT results (data
95 not shown).

96 We then assessed the failure rate of GSS prepared using urethral and meatal
97 swabs, with failure defined as an absence of cellular material on microscopy (*i.e.*
98 “quantity of cells not sufficient” [QNS]). The GSS QNS rate of meatal swabs was
99 significantly higher compared with urethral swabs (45% vs. 3%, $P < 0.0001$). Despite no
100 difference in the frequency of signs or symptoms of urethritis between the two groups,
101 meatal swabs were associated with significantly lower number of GSS with PMN
102 between 2-5 (3% vs. 33%, $P = 0.0011$, Table 1) than urethral swabs, which is reflected
103 in the high GSS failure rate.

104 To determine if the increased QNS rate in meatal GSS was associated with less cellular
105 content, we stratified the SAC_{CT} results by QNS status. As shown in Figure 1A, Gram
106 stains identified as QNS were associated with a higher meatal swab SAC_{CT} (mean 27.4
107 vs 24.1, $P = 0.0002$), indicating that the meatal swabs used to prepare the GSS
108 contained less cellular material.

109 Given our finding that meatal swabs are associated with significantly less cellular
110 material and a higher GSS failure rate compared with urethral swabs, we were
111 interested in establishing how meatal swabs performed when sampling discharge. As
112 shown in Figure 1B, in the absence of discharge, meatal swabs collected significantly
113 less cellular material than urethral swabs (mean SAC_{CT} 27.0 vs 24.4, $P = 0.0003$). If
114 discharge was present, the meatal swab collected significantly more cellular material
115 than a meatal swab from men without discharge (mean SAC_{CT} 23.7 vs 27.0, $P =$

116 0.0004). Further, in the setting of visible discharge, no difference in the cellular content
117 collected comparing the meatal or urethral swabs was identified (mean SAC_{CT} 23.7 vs
118 23.3, $P = 0.4789$). Evaluating the GSS failure rate of swabs in the presence or absence
119 of discharge, the highest QNS rates were seen in meatal swabs from men without
120 discharge compared to meatal swabs from men with discharge (68% vs 15%, $P =$
121 0.0022, Figure 1C) and compared to urethral swabs in men without discharge (68% vs
122 6%, $P < 0.0001$, Figure 1C). In the presence of discharge, meatal swabs were
123 associated with a slight non-significant increase in the QNS rate compared to urethral
124 swabs (15% vs 0%, $P = 0.1373$, Figure 1C).

125 We then compared the diagnoses (using both Xpert® and GSS results) of all
126 QNS results to assess the number of NG or NGU diagnoses that could have been
127 missed by GSS failures. Of the 16 QNS GSS results, none were NG diagnoses, but
128 three NGU diagnoses (one CT diagnosed by NAAT and two non-CT NGU [defined by
129 discharge on exam]) were included, which highlights that GSS failures from meatal
130 swabs could delay the time to effective treatment (*i.e.*, missed opportunity to evaluate
131 PMNs in point-of-care testing) in the absence of discharge (data not shown).

132

133 Discussion

134 Evaluation by GSS of urethral secretions from a urethral or meatal swab remains
135 the test-of-choice for rapid diagnosis of NG or NGU in men with urethritis symptoms,
136 though no preferred swab type is recommended by the 2015 STD Treatment Guidelines
137 for collecting urethral secretions (7). In this study, we compared meatal and urethral
138 swabs from men and used the Cepheid Xpert® CT/NG SAC_{CT} to measure the

139 difference in the amount of swab cellular material and determine the failure rate of GSS
140 testing for each swab type. Not surprisingly, we found that meatal swabs collected
141 significantly less cellular material, compared with urethral swabs, in the absence of
142 urethral discharge. Moreover, meatal swabs were associated with a high GSS failure
143 rate, eliciting a 12-fold increase in QNS GSS rates, compared with urethral swabs, in
144 men without discharge. In the presence of urethral discharge, there was no difference in
145 the cellular content or GSS failure rate between the swab types, indicating that the
146 collection quality of meatal swabs is highly susceptible to the sampling surface area of
147 the urethral meatus and/or may be collecting increased numbers of non-intact cells;
148 limitations that may be overcome by urethral sampling. Additionally, the limitations of
149 meatal swabs may only significantly affect Gram stain testing, which is dependent on
150 the collection of intact cells to evaluate for the presence of GNID and PMNs, whereas
151 NAAT testing is much more sensitive and less likely to be significantly influenced by
152 changes in the swab cellular content. In fact, NAAT testing comparing urine and either
153 swab type demonstrated a 100% concordance rate for the diagnosis of CT and/or NG in
154 our study, despite the meatal swabs containing less cellular material.

155 Although the NAAT performance of self-collected meatal swabs appears
156 comparable to clinician-collected urethral swabs (5) and superior to urine for diagnosis
157 of CT, NG, and trichomonas (6), no study has compared the performance of meatal
158 swabs to urethral swabs for Gram stain point-of-care testing to evaluate urethritis (7). To
159 our knowledge, our study is the first to demonstrate that in the absence of urethral
160 discharge, meatal swabs are inferior to urethral swabs at collecting cellular material,
161 which increases the failure rate of GSS testing. In addition, although there was no

162 difference in the signs or symptoms between the swab groups, meatal swabs were
163 associated with significantly fewer numbers of GSS with PMN 2-5 (Table 1), suggesting
164 that under-diagnosis of NGU cases may also result when meatal swabs are used for
165 GSS testing.

166 Our findings have important implications for the use of meatal swabs to collect
167 urethral secretions, a practice that may be increasing given that meatal swabs have
168 several advantages over urethral swabs for NAAT testing. Meatal swabs are likely
169 better tolerated than urethral swabs as demonstrated in a comparison study of men who
170 received both a urethral and meatal swab, which found that 76% of men preferred the
171 meatal swab (4). Another study attempted to quantify the discomfort associated with
172 urethral swab collection using a visual analogue pain scale and found that collection of
173 a urethral swab using standard technique (inserting a swab 2 cm into the distal urethra)
174 elicited a median pain score of 52 mm (on a 100-mm pain scale), which correlated with
175 moderate (>30 mm) to severe (>54 mm) pain (10). Furthermore, multiple studies have
176 identified discomfort from urethral swabbing as a major factor that causes men to delay
177 seeking health care (11-13) and participants in adolescent focus groups expressed
178 strong negative emotions when asked about the urethral swab testing process (12, 13).
179 In addition, the ease of obtaining a meatal swab facilitates self-collection for NAAT
180 testing, allowing patients to self-collect specimens at home or in a clinical setting where
181 regular interval screening of asymptomatic, high-risk men is performed. Given that 77%
182 of men preferred self-collection of a meatal swab to provider-collection (14) and over
183 90% of men report self-collection of a penile-meatal swab as “easy” or “very easy” (5,
184 14), it is likely that NAAT testing of self-collected meatal swabs may play an important

185 future role in screening asymptomatic men in busy, high-flow clinical settings or non-
186 clinical settings.

187 Our study has several limitations. We could not evaluate the performance of
188 meatal swabs versus urethral swabs for the diagnosis of NGU or NG by Gram stain as
189 our study wasn't powered to evaluate that outcome. Also, the majority of men in our
190 study were African American, which is reflective of demographic characteristics of
191 clients attending the STD clinic and our results may not be generalizable to other, non-
192 STD Clinic populations. Additionally, the Gram stains were prepared by multiple
193 clinicians, all highly proficient at preparing and reading Gram stain smears. Although no
194 difference in either swab cellular content or the rate of QNS GSS results was associated
195 with any individual clinician (data not shown), we cannot rule out the possibility that
196 differences in provider collection techniques may have influenced our results.

197 In conclusion, in the absence of visible urethral discharge, the use of meatal
198 swabs for point-of-care diagnosis of NG or NGU by GSS testing was associated with a
199 significant decrease in swab cellular content and an increase in GSS failure rates,
200 compared to urethral swabs. Therefore, in the absence of discharge, meatal swabs
201 should be avoided when considering point-of-care testing for NG or NGU in men.

202

203 **Materials and Methods**

204 We recruited Men \geq 19 years old from the Jefferson County Department of
205 Health (JCDH) STD clinic in Birmingham, Alabama, excluding only men who had voided
206 within the past 60 minutes and those who had received antibiotics with CT/NG activity
207 within the past 30 days. After informed consent was obtained, a directed physical exam

208 was performed for the presence or absence of discharge and men were alternately
209 assigned to receive either a urethral or meatal swab for specimen collection using a
210 dacron swab. To collect a urethral swab, highly experienced clinicians inserted the swab
211 2cm into the urethra and rotated during insertion and extraction. For meatal swab
212 specimen collection, the swab was positioned perpendicular to the urethral meatus and
213 rolled back and forth several times across the meatus for 5 seconds. Following
214 specimen collection, swabs were immediately rolled onto the surface of a glass
215 microscope slide in preparation for Gram stain of urethral secretions and then placed in
216 an Xpert® CT/NG (Cepheid, Sunnyvale, CA) specimen transport container and stored
217 at 4°C. Following meatal or urethral swab collection, an initial void urine specimen was
218 collected for CT and NG testing.

219 Gram stain of the smear was performed using standard laboratory procedure and
220 all smears were read by a highly-experienced clinician (JRS), blinded to the physical
221 exam findings and Xpert® CT/NG assay results. After identifying an area of high
222 cellularity using low magnification, the presence or absence of Gram negative
223 intracellular diplococci (GNID) and average number of leukocytes from five contiguous
224 high power fields were determined and recorded using high magnification with oil
225 immersion (1000x). GSS were labeled “Quantity cells not sufficient” (QNS) if no cells
226 were identified by scanning either low or high power magnification. NGU was diagnosed
227 if no GNID were seen and either (1) a discharge was present on exam or (2) ≥ 2 PMN's
228 per HPF were seen by GSS testing.

229 Xpert® CT/NG testing was performed on both urethral and meatal swab
230 specimens and post-swab urine specimens within 7 days of collection as described in

231 the package insert. For quantification of specimen cellularity, the Xpert® CT/NG sample
232 adequacy control crossing threshold (SAC_{CT}) was used. We also tested the
233 performance of each swab type to prepare a GSS by determining the failure rate of
234 GSS testing, defined as lacking sufficient number of cells to evaluate the specimen.
235 This study was approved by the institutional review board of the University of Alabama
236 at Birmingham and the research review committee at JCDH. Bivariate comparisons
237 were evaluated using an unpaired T-test while Fisher's Exact or Chi-square tests were
238 used to test differences between groups. Significance was reported as $P < 0.05$ using
239 Prism software (v7.0b; Graphpad Software, Inc., San Diego, CA).

240

241

242 Acknowledgments:

243 The authors are grateful to Paula Dixon and Austin Culver for assistance with specimen
244 processing and testing.

245

246

247 Funding:

248 S.J.J. has no conflicts of interest. J.R.S. has received honoraria, consulting fees, or
249 research support from Cepheid, BD Diagnostics, LabCorp, and Hologic. K.J.A. has no
250 conflicts of interest. B.V.D.P. has received honoraria, consulting fees, or research
251 support from the following sponsors: Abbott Molecular Diagnostics, Atlas Genetics, BD
252 Diagnostics, Beckman Coulter, Great Basin, Scientific, Cepheid, Hologic, Rheonix, and
253 Roche Diagnostics. E.W.H. has received honoraria, research support, or consulting fees

254 from Cepheid, BD Diagnostics, Gen-Probe Hologic, Roche Diagnostics, and Cempra
255 Pharmaceuticals. Reagents and test kits for this study were supplied by Cepheid
256 (Sunnyvale, CA). This work was funded from the National Institute of Allergy and
257 Infectious Diseases of the National Institutes of Health Sexually Transmitted Infection
258 Cooperative Research Center grant U19AI113212 (E.W.H., PI).

259

260 Figure Legend:

261 **FIGURE 1.** Comparison of cellular content and percent QNS in meatal versus urethral
262 swabs in the presence or absence of urethral discharge. (A) Meatal swabs that resulted
263 QNS by GSS are associated with a significantly higher SAC_{CT}, compared with
264 satisfactory GSS, indicating they contain less cellular material. (B) In the absence of
265 discharge, meatal swabs have a significantly higher SAC_{CT} compared to urethral swabs
266 from men without discharge or meatal swabs from men with discharge. In the presence
267 of discharge, there was no difference in the SAC_{CT} between meatal or urethral swabs.
268 (C) In the absence of discharge, meatal swabs were significantly more likely to result
269 QNS by GSS compared to urethral swabs from men without discharge or meatal swabs
270 from men with discharge. In the presence of discharge, there was no difference
271 between the swab types. Horizontal lines and whiskers denote the mean and 95% CI,
272 respectively. SAC_{CT} values were obtained from the Xpert® CT/NG assay.

273

274

275

276 References:

- 277 1. **Centers for Disease Control and Prevention.** 2015. Sexually transmitted
278 disease surveillance 2014. Atlanta: U.S. Department of Health and Human
279 Services.
- 280 2. **Moi H, Blee K, Horner PJ.** 2015. Management of non-gonococcal urethritis. BMC
281 Infectious Diseases **15**:294.
- 282 3. **Papp JR, Schachter J, Gaydos C, Van Der Pol B.** 2014. Recommendations for
283 the laboratory-based detection of *Chlamydia trachomatis* and *Neisseria*
284 *gonorrhoeae*--2014. MMWR **63**:1–19.
- 285 4. **Lamba H, Davies J, Murphy S, Shafi M.** 2001. Detection of chlamydia on meatal
286 swabs. Sex Transm Infect **77**:224–224.
- 287 5. **Dize L, Barnes P, Barnes M, Hsieh Y-H, Marsiglia V, Duncan D, Hardick J,**
288 **Gaydos CA.** 2016. Performance of self-collected penile-meatal swabs compared
289 to clinician-collected urethral swabs for the detection of *Chlamydia trachomatis*,
290 *Neisseria gonorrhoeae*, *Trichomonas vaginalis*, and *Mycoplasma genitalium* by
291 nucleic acid amplification assays. Diagnostic Microbiol and Infect Dis **86**:131–135.
- 292 6. **Dize L, Agreda P, Quinn N, Barnes MR, Hsieh Y-H, Gaydos CA.** 2013.
293 Comparison of self-obtained penile-meatal swabs to urine for the detection of *C.*
294 *trachomatis*, *N. gonorrhoeae* and *T. vaginalis*. Sex Transm Infect **89**:305–307.
- 295 7. **Workowski KA, Bolan GA, Papp JR.** 2015. Sexually transmitted diseases
296 treatment guidelines, 2015. MMWR.
- 297 8. **Jordan SJ, Van Der Pol B, Hook EW.** 2017. Utilization of the Cepheid Xpert®
298 CT/NG sample adequacy control to determine the influence of the urethral swab

- 299 on cellular content in post-swab versus pre-swab urine. *Sex Transm Dis* **44**:67–
300 68.
- 301 9. **C Bristow C, Adachi K, Nielsen-Saines K, Ank B, Morgado MG, Watts H,**
302 **Veloso VG, Pilotto JH, Joao EC, Klausner JD.** 2014. Characteristics of the
303 sample adequacy control (SAC) in the Cepheid Xpert® CT/NG assay in female
304 urine specimens. *JMEN* **1**:1–5.
- 305 10. **Apoola A, Herrero-Diaz M, FitzHugh E, Rajakumar R, Fakis A, Oakden J.**
306 2011. A randomised controlled trial to assess pain with urethral swabs. *Sex*
307 *Transm Infect* **87**:110–113.
- 308 11. **Armstrong B.** 2003. The Young Men's Clinic: addressing men's reproductive
309 health and responsibilities. *Perspect Sex Reprod Health* **35**:220–225.
- 310 12. **Blake DR, Kearney MH, Oakes JM, Druker SK, Bibace R.** 2003. Improving
311 participation in chlamydia screening programs: perspectives of high-risk youth.
312 *Arch Pediatr Adolesc Med* **157**:523–529.
- 313 13. **Tilson EC, Sanchez V, Ford CL, Smurzynski M, Leone PA, Fox KK, Irwin K,**
314 **Miller WC.** 2004. Barriers to asymptomatic screening and other STD services for
315 adolescents and young adults: focus group discussions. *BMC Public Health* **4**:21.
- 316 14. **Chai SJ, Aumakhan B, Barnes M, Jett-Goheen M, Quinn N, Agreda P, Whittle**
317 **P, Hogan T, Jenkins WD, Rietmeijer CA, Gaydos CA.** 2010. Internet-based
318 screening for sexually transmitted infections to reach nonclinic populations in the
319 community: risk factors for infection in men. *Sex Transm Dis* **37**:756–763.
- 320

Characteristic	Total (n = 66)	Urethral Swab (n = 33)	Meatal Swab (n = 33)	P-value
Age, mean (range)	29 (20 – 69)	29 (20 – 60)	29 (20 – 69)	0.9720
Race, n (%)				0.2920
AA	56 (89%)	28 (85%)	28 (93%) ^a	
Caucasian	7 (11%)	5 (15%)	2 (7%) ^a	
Other	0	0	0 ^a	
Prior STI	43 (65%)	25 (76%)	18 (55%)	0.0724
Partners, n last 30 days	1.6 (0 – 7)	1.8 (0 – 7)	1.5 (0 – 4)	0.4472
Mean (range)				
Discharge Present, n (%)	27 (41%)	14 (42%)	13 (39%)	0.8040
Gram Stain, n (%)				
QNS	16 (24%)	1 (3%)	15 (45%)	< 0.0001
GNID positive	9 (14%)	3 (9%)	6 (18%)	0.2891
GNID negative				
PMN < 2	9 (14%)	7 (21%)	2 (6%)	0.0749
PMN 2 – 5	12 (18%)	11 (33%)	1 (3%)	0.0011
PMN ≥ 5	20 (30%)	11 (33%)	9 (27%)	0.5989
Xpert® Swab Results				
Neg	46 (70%)	25 (76%)	21 (64%)	0.2912
CT+	8 (12%)	2 (6%)	6 (18%)	0.1556
CT+NG+	4 (6%)	2 (6%)	2 (6%)	> 0.999
NG+	8 (12%)	4 (12%)	4 (12%)	> 0.999
SAC _{CT} , mean (range)	24.8 (21 – 33)	23.9 (21 – 27)	25.6 (21 – 33)	0.0026
Urine SAC _{CT} , mean (range)	26.1 (19 – 32)	26.3 (19 – 32)	25.9 (19 – 32)	0.6147

Abbreviations: AA, African American; STI, sexually transmitted infection; GNID, Gram-negative intracellular diplococci; PMN, polymorphonuclear neutrophil; QNS, quantity cells not sufficient; CT, chlamydia; NG, gonorrhea

^a3 missing

Significance evaluated using Fisher's Exact test or T-test, as appropriate.

