Pilot clinical study to assess caries lesion activity using quantitative light-induced fluorescence during dehydration

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Abstract. This study aimed to evaluate the ability of quantitative light-induced fluorescence (QLF) to assess caries lesion activity using visual examination (VE) as the gold standard. Twenty-four visible white spot lesions on buccal surfaces were examined from 23 children, ages 9 to 14 years. At baseline, the surface was hydrated with water, and thereafter, it was dehydrated with continuous compressed air during image acquisition. QLF images were acquired at 0 (baseline), 5, and 15 s. QLF variables [QLFV: fluorescence loss (∆F), lesion size (S), ΔQ: ΔF × S] was recorded. Changes-in-QLFV per second (ΔQLFV) were determined: ΔQLFV = (QLFV - QLFVBaseline) / N, where N indicates dehydration time. One experienced dentist conducted VE independently using a dental unit’s light, compressed air, and explorer. QLFV and ΔQLFV of the active group (n = 11) were compared with those of the inactive group (n = 13) using two-sample t-tests. As the surface was dehydrated, S and ΔQ values of the active group increased, whereas QLFV of the inactive group showed only a small change. ΔQLFV of the active group were larger than those of the inactive group; however, the difference did not reach statistical significance (p > 0.11). Within the limitations of this study, QLF data indicated increments for lesions designated as active and minimal change for lesions defined as inactive. © 2017 Society of Photo-Optical Instrumentation Engineers (SPIE) [DOI: 10.1117/1.JBO.22.3.035005]

Keywords: clinical study; caries lesion activity; diagnostics; fluorescence.

1 Introduction

Dental caries is a dynamic process during which demineralization and remineralization cycles take place continuously. When the demineralization process continues, it increases surface porosity and creates white spot lesions due to increasing refractive index. Leaving these lesions untreated can lead to cavitation. By detecting the early stage of dental caries, such as white spot lesions, it allows to arrest the demineralization process before surgical treatment is considered and even to gain net mineral. In order to achieve this, early detection and caries activity assessment are critical. In addition to visual methods, several technology-based methods have been introduced.

Quantitative laser- and light-induced fluorescence (QLF) techniques are among the most widely studied methods for detection of early caries. They have been used widely in various in vitro, in situ, and in vivo studies. A study showed both laser and light fluorescence to be useful in detecting and quantifying white spot lesions. QLF has been used in a controlled clinical trial to show significant differences between fluoride dentifrice treatment regimens in as little as 3 months. This suggests that the use of QLF in clinical trials may reduce the time necessary to evaluate new therapeutic agents, saving both time and expense. The main advantage of this technique is that it can be used to quantify changes in carious lesions over time.

Near-infrared (NIR) imaging has been shown to detect and quantify dental caries. Also, both thermal and NIR reflectance imaging have been found to be suitable for the detection of remineralization of simulated caries lesions. The use of optical coherence tomography (OCT) for caries detection and quantification has also shown promising results. Another study reported that polarization-sensitive OCT (PS-OCT) and dehydration rate measurements with NIR reflectance and thermal imaging methods were ideally suited for nondestructive, noninvasive, and quantitative measurement of lesion activity.

Based on chemical analysis and histopathological observations, the initial stage of caries development is characterized by the opening of the intercrystalline spaces without the destruction of the surface and subsequent creation of microchannels. These microchannels are found to be about 0.5 to 1.5 μm in width in artificial lesions and range from 0.2 to 1.0 μm in width in early natural enamel lesions. When white spot lesions (the early stage of demineralized enamel) are hydrated/wet, microchannels in lesions are filled with fluid/water. The presence or absence of water in microchannels affects the amount of light scatter and internal reflection. The difference between the refractive indices of water (1.33) and enamel crystal (1.62) is minimal. Because this difference is small, when the surface is wet (hydrated), there is less scatter and the light path is longer than...
when the surface is dry (dehydrated). Light absorption and fluo-
rescence remitted per volume is much greater in the hydrated
white spot lesions and fluorescence is stronger. This may be
the reason white spot lesions may not be readily noticeable when
the surface is wet (hydrated). However, when the lesions are
dehydrated/dry, air fills the porous areas (intercrystalline spaces).
Air has a lower refractive index (nearly 1.0). Differences
between the refractive indices of intercrystalline space (air) and
enamel crystal increase, hence the amount of light scattering
increases. Light absorption and fluorescence remitted per vol-
ume is much smaller in the dehydrated white spot lesions and
fluorescence is weaker. This may be the reason the white spot
lesions are more readily recognized when the surface is dry
(dehydrated).

A caries lesion that is progressing (continuing to demineral-
ize) is described as an active caries lesion. A lesion that has
stopped further progression (stagnant/renmineralized) is referred
to as an inactive or arrested caries lesion. There are several
reports on the characteristics of caries lesion activity in the
literature. A high reliability for assessment of caries lesion
activity was shown with criteria based on observations of sur-
face appearance and texture. For intact surfaces, active caries
lesions were described as presenting a whitish/yellowish opaque
enamel surface with a loss of luster and a rough texture when the
tip of the probe is moved gently across the surface, whereas
inactive caries lesions were described as presenting a shiny
enamel surface and a hard and smooth texture. By definition,
active caries lesions must have microchannels (increased poros-
ity) that allow biofilm-generated acids to penetrate into the
lesion body for caries to progress. On the other hand, inactive
lesions should have smaller and/or fewer microchannels, indi-
cating the caries process has been reversed or arrested. When
lesions are wet, microchannels in active white spot lesions are
filled with water and the amount of water is larger than in inac-
tive white spot lesions. Inactive white spot lesions have smaller
microchannels relative to active lesions, therefore, they have
less water/fluuid. As the size and/or number of microchannels
increases, the surface porosity increases, thus water/fluuid inside
the intercrystalline spaces evaporates faster. Therefore, the cen-
tral hypothesis of this study was that the rate and amount of
vaporization (evaporation) of fluid in the lesion body during
dehydration as measured by QLF indicates caries lesion activity.
The change-in-QLF variables per second ($\Delta QLF_v$) during dehy-
dration would show relatively large values due to the presence
of wider/longer microchannels and larger amounts of water in
the lesion.

Ando et al. demonstrated in extracted human teeth that dur-
ing the first few seconds of dehydration by continuous com-
pressed air, change-in-QLF variables per second ($\Delta QLF_v$) values for active lesions were larger than those of inactive
lesions. This suggests that $\Delta QLF_v$ during the first few seconds
of dehydration by continuous compressed air may be able to
differentiate between active (demineralizing) and inactive
(renmineralized) caries lesions at the time of examination. To
the authors’ best knowledge, QLF during dehydration had not
been tested clinically to assess caries lesion activity. Therefore,
the objective of this clinical pilot study was to determine if
changes-in-QLF variables during dehydration could be used
to assess caries lesion activity of noncavitated caries lesions
(white spot lesions) on the buccal surface in permanent teeth of
children in vivo. Accomplishing the specific objective of this
clinical pilot study would be of substantial significance
for dental public health. Particularly, this study would evaluate
the use of the objective and quantitative QLF during dehydration
technique for determination of caries lesion activity at the time
of examination. Using an objective and quantitative method
(QLF with dehydration) could potentially improve clinical deci-

dion-making for more effective caries management.

2 Subjects and Methods

This study protocol was approved by the Indiana University
Institutional Review Board (IRB #0105-10). The subject’s
parents completed an informed consent medical/dental history
and children signed an assent. Upon review and subsequent
acceptance into the study, a visual examination (VE) was per-
fomed on all of the teeth in the mouth. Subjects who had at least
one white spot lesion, no severe cavitated caries lesions, no
moderate to severe periodontal diseases, and no orthodontic
appliances were included. Twenty-three children, ages 9 to
14 years, presented 24 visible white spot lesions on the buccal
surface of permanent teeth at the gingival margin for exami-
nation. One experienced dentist, trained and calibrated in the
International Caries Detection and Assessment System
(ICDAS) and Nyvad’s criteria, independently conducted the
VE to assess caries lesion activity using a dental unit’s
light and compressed air, with an explorer used only to check
surface structure. The examiner utilized the best knowledge
from both criteria to assess caries lesion activity using reflection
and tactile sensation. Active caries lesions were defined as pres-
centing a whitish/yellowish opaque enamel surface with a loss
of luster and a rough texture when the tip of the explorer was
moved gently across the surface. Inactive caries lesions were
defined as presenting a shiny enamel surface and a hard and
smooth texture when the tip of the explorer was moved gently
across the surface. In order to reduce/eliminate bias, a separate
examiner performed an examination using QLF with the dehy-
dration technique.

2.1 Quantitative Light-Induced Fluorescence
Examination

Three consecutive fluorescence images per surface were
acquired at 0 (baseline), 5, and 15 s with the quantitative light-
induced fluorescence system (QLFPatient 3.0.0.35, Inspektor
Research Systems B.V., The Netherlands). These intervals
between image acquisition were based on the previous in
vitro study. The state of hydration of the surfaces was stan-
dardized by placing a wet cotton pellet, which was soaked in dis-
tilled water, on the buccal surface for 60 s. The QLF camera
handpiece was maintained in a position that provided the best
illumination of the buccal surface. The subject bit and held a
short piece of wooden tongue depressor and the QLF camera
handpiece was placed against this to secure the position.
During image acquisition while the examiner maintained the
QLF camera position, the lesions were dehydrated with contin-
uous compressed air by an assistant. After the image acquisition,
QLF variables ($QLF_v$) of fluorescence loss [$\Delta F (%)$, lesion size
[$S$ (mm$^2$)], and $Q_\lambda$: $\Delta F \times S$ ($\% \times mm^2$)] were determined with
proprietary QLF software (QLF 2.00 g, Inspektor Research
Systems B.V., The Netherlands). The details of image analysis
with QLF software were described previously. Briefly, a lesion
threshold <95% of reconstructed sound fluorescence radiance
level was considered to be a caries lesion.
2.2 Data Analyses

Changes-in-QLF variables per second \([\Delta \text{QLF}_{VN}, \Delta F_{VN} (\%/s), \Delta S_{VN} (\text{mm}^2/s), \Delta \text{Q}_{VN} (\% \times \text{mm}^2/s)]\) were determined using the following equation: \(\frac{\text{QLF}_{VN}(\text{baseline})}{N}\), where \(N\) indicated dehydration/image acquisition time.

QLF\(_V\) and \(\Delta \text{QLF}_{VN}\) values were divided into active and inactive white spot lesion groups based on the VE. The averages and standard errors of QLF\(_V\) and \(\Delta \text{QLF}_{VN}\) values of each dehydration (image acquisition) time were calculated for the active and inactive groups. Comparisons between active and inactive white spot lesions at each dehydration time were performed using 2-sample \(t\)-tests, and comparisons of the measurements during dehydration were made using repeated measures ANOVA. A 5% significance level was used for all tests. Statistical analyses were performed using SAS version 9.1 (SAS Institute Inc., Cary, North Carolina).

### 3 Results

Eleven out of 24 sites were designated as active status according to VE, and 13 sites were designated as inactive status. Figure 1 shows examples of fluorescence images of both active and inactive groups at 0, 5, and 15 s of dehydration. Caries lesions appeared darker than sound area. As dehydration time increased, active caries lesion got darker. On the other hand, inactive caries lesion did not get darker. Figure 2 displays the average and standard error of fluorescence loss. Although there were no statistically significant differences between groups (0 s: \(p = 0.16\), 5 s: \(p = 0.09\), 15 s: \(p = 0.14\)) or among dehydration times (active: 0 s versus 5 s: \(p = 0.92\), 0 s versus 15 s: \(p = 0.59\), 5 s versus 15 s: \(p = 0.53\); inactive: 0 s versus 5 s: \(p = 0.44\), 0 s versus 15 s: \(p = 0.82\), 5 s versus 15 s: \(p = 0.35\)), more loss of fluorescence was observed in the active lesions. However, dehydration for either 5 or 15 s did not alter the amount of fluorescence loss for either type of lesion.

Figure 3 similarly presents the effect of dehydration on the size of the lesions as assessed by QLF. While again there were no statistically significant differences between groups (0 s: \(p = 0.14\), 5 s: \(p = 0.07\), 15 s: \(p = 0.07\)) or among dehydration times (active: 0 s versus 5 s: \(p = 0.22\), 0 s versus 15 s: \(p = 0.08\), 5 s versus 15 s: \(p = 0.25\); inactive: 0 s versus 5 s: \(p = 1.00\), 0 s versus 15 s: \(p = 0.40\), 5 s versus 15 s: \(p = 0.40\)), active lesions were observed to be larger in size (\(\text{mm}^2\)). Dehydration did not have obvious influence on apparent lesion size for the inactive lesions, but an apparent increase in lesion size was observed for active lesions following dehydration.

Figure 4 shows the impact of dehydration on the volume (\(\Delta \text{Q}\)) of active and inactive lesions. There were no statistically significant differences between groups (0 s: \(p = 0.22\), 5 s: \(p = 0.11\), 15 s: \(p = 0.12\)) or among dehydration times (active: 0 s versus 5 s: \(p = 0.41\), 0 s versus 15 s: \(p = 0.11\), 5 s versus 15 s: \(p = 0.22\); inactive: 0 s versus 5 s: \(p = 0.57\), 0 s versus 15 s: \(p = 1.00\), 5 s versus 15 s: \(p = 0.56\)). Influenced by the results for lesion size, dehydration had little effect on the volume (\(\Delta \text{Q}\)) of inactive lesions but was increased slightly in active

<table>
<thead>
<tr>
<th>Activity status</th>
<th>Dehydration / image acquisition time (s)</th>
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<tbody>
<tr>
<td></td>
<td>0 (Baseline)</td>
</tr>
<tr>
<td>Active</td>
<td><img src="image1.png" alt="Image" /></td>
</tr>
<tr>
<td>Inactive</td>
<td><img src="image4.png" alt="Image" /></td>
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**Fig. 1** Example of fluorescence images of both active and inactive groups at 0 (baseline), 5, and 15 s of dehydration.

**Fig. 2** Average and standard error of fluorescence loss for active and inactive groups. There were no significant differences between groups or among dehydration times \((p > 0.05)\).

**Fig. 3** Average and standard error of size of lesion for active and inactive groups. There were no significant differences between groups or among dehydration times \((p > 0.05)\).

**Fig. 4** Average and standard error of \(\Delta \text{Q}\) for active and inactive groups. There were no significant differences between groups or among dehydration times \((p > 0.05)\).
lesions. This was in contrast to the QLFV of inactive lesions, which showed minimal change during dehydration.

Table 1 presents the average and standard error of the changes-in-QLF variables per second (ΔQLFV) for the active and inactive groups. Although ΔQLFV values of active lesions were larger than those of inactive lesions, the differences between groups did not reach statistical significance (5 s ΔFV: p = 0.49, 5 s ΔSV: p = 0.11, 5 s ΔQV: p = 0.12, 15 s ΔFV: p = 0.78, 15 s ΔSV: p = 0.11, 15 s ΔQV: p = 0.12).

To explore how the changes-fared as a diagnostic test, a cut-off value for the change-in-ΔQ per second (ΔQV) was selected at ΔQV = 1.2. With this cut-off, sensitivity was 100%, specificity was 62%, positive predictive value was 69%, and negative predictive value was 100%. Given the small sample size in this study, this may not be the optimal cut-off; however, it does show that the method has relevance as a possible method to identify activity.

4 Discussion

In the postfluoride era, progression of enamel caries is slower, and it takes longer before the stage of irreversible destruction of the tooth is reached (cavitation).25 With new assessment methods, such as QLF, caries detection is possible at an earlier stage than could be performed previously. This is very important for carrying out timely and appropriate preventive measures. However, as caries is an irreversible and reversible process in its early stages, identification of only the severity of caries lesions, such as lesion size and depth, is insufficient. We know some of these lesions will be active and, therefore, in need of some form of intervention, while others may be a scar of past damage (inactive/arrested) and, therefore, will not require any intervention. In order to select an appropriate treatment modality and aid in the clinical decision-making process, caries diagnosis and assessment of caries lesion activity are of critical importance. Incorrect diagnosis can easily result in incorrect treatment decisions, particularly with respect to irreversible treatments (i.e., restorations).

The key feature that allows QLF imaging to differentiate between active and inactive lesions is based on their differences in the surface structure, specifically the porosity of the surface layer. As mentioned previously, active lesions may present a porous surface layer and inactive lesions may have a relatively nonporous surface layer. The porosity depends on the number and size of microchannels. A study using scanning electron microscopy indicated that microchannels developed at caries initiation and increased in size with continued demineralization.26 As the size and/or number of microchannels increases, the surface porosity increases. On the other hand, inactive lesions should have fewer and narrower microchannels. This implies that the surface porosity decreases. For active caries lesions, based on our hypothesis, the QLFV and ΔQLFV during dehydration would show relatively large values, due to the presence of wider/longer microchannels, and larger amounts of water in the lesion. On the other hand, for inactive caries lesions, QLFV and ΔQLFV would show much lower values during dehydration, due to the presence of smaller and fewer microchannels, and thus a negligible amount of water in the lesion.

In this pilot clinical study, although not reaching statistical significance, the results appear to support our hypothesis by indicating there was a trend for ΔQ of active lesions to increase as the surface was dehydrated, leading to higher change-in-ΔQ (ΔQV) for active lesions than for inactive lesions. A poststudy power analysis was performed to evaluate the role of the small sample size on the lack of statistical significance for this study. The observed differences in ΔQV were 0.45 and 0.25 at 5 and 15 s of dehydration, respectively. A study with n = 42 per group would have 80% power at a 5% significance level to detect the ΔQV differences we observed. Therefore, this pilot clinical study suggests that QLF imaging of caries lesions undergoing dehydration can be used to differentiate caries lesion activity. This is advantageous as it can be done at the time of examination, and information can be obtained in a single appointment. However, further investigations are needed to determine the ΔQV threshold to differentiate active and inactive caries lesions.

There were two other studies of fluorescence imaging with dehydration that may be relevant to this study. Al-Khateeb et al.27 indicated that dehydration data could serve as a separate method for analyzing pore distribution properties in lesions to differentiate history of demineralization or fluoride treatment. Van der Veen et al.28 demonstrated the potential to use fluorescence imaging with dehydration to determine caries lesion activity. For both studies, dehydration was done by allowing the specimens to dry at room temperature rather than using controlled compressed air. These studies may not be directly applicable to a clinical setting. Clinically another study has demonstrated that lesions which progress to cavitation (active) present faster changes in QLFV than inactive lesions.3 Another clinical study, using a different laser fluorescence device (LF: this consists of a 655-nm emitted diode laser) with occlusal caries, demonstrated that after air drying for 3 and 15 s, LF values of active caries, in which caries activity was determined by VE, presented higher than those of inactive caries.29

There were also studies regarding the dehydration effect using OCT. For one study, dehydration was induced by the specimens in air at room temperature for 30 min. The results showed that hydration state affects the reflectivity of demineralized porous enamel, and the effect can be potentially used for assessment of early enamel lesion using OCT.30 For another study, dehydration was produced in a controlled environment for 30 s. The air pressure was set to 15 psi, and the computer-controlled air nozzle was positioned 2 cm away from the sample.31 Although bovine dentin and extracted human root caries were used,
thermal imaging and PS-OCT may be ideally suited for the non-destructive root caries lesion activity during a clinical examination. Therefore, these studies and this study suggested that the state of hydration in the lesion body can be used to quantify severity of dental caries and caries lesion activity.

For this proof-of-concept study, we intended to demonstrate that QLF data could indicate increments for lesions designated as active and minimal change for lesions defined as inactive. Hence, the design of this study was small sample size and cross sectional proof-of-concept pilot study. VE was employed as the gold standard. The examiner for VE in this study is trained and calibrated with ICDASII and Nyvad’s criteria and is expert in caries detection and diagnosis. Previous studies demonstrated that both criteria could predict lesion depth and assess caries lesion activity. Although not reaching statistical significance, this small, pilot clinical study provided preliminary information to establish the use of the QLF method. Further clinical investigations to establish threshold or cut-off values to distinguish among the lesion stages and a larger sample size for clinical application will be necessary.

5 Conclusion

The results of this small, pilot clinical study suggest that QLF data, especially ΔQ, indicated slight increments for lesions designated as active and minute changes for lesions defined as inactive. Furthermore, change-in-Q per second (ΔQΔt) of active lesions was greater than that of inactive lesions. Therefore, QLF during dehydration has the potential to assess caries lesion activity of noncavitated caries lesions (white spot lesions) on smooth surface in vivo at the time of examination.

Disclosures

George K. Stookey is the owner of Therametric Technologies, Inc.; he has an interest in the use of fluorescence for early caries detection, but no financial interest in QLF. The other authors declare no conflict of interest.

Acknowledgments

This study was supported by the 21st Century Research and Technology Fund from the State of Indiana, USA. The funder had no role in study design, data collection and analysis, decision to publish, or preparation of the paper.

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Journal of Biomedical Optics 035005-5 March 2017 • Vol. 22(3)

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