

1  
2  
3  
4 **Title**  
5

6  
7 Effect of different endodontic regeneration protocols on wettability, roughness and  
8 chemical composition of surface dentin  
9

10  
11  
12  
13 **Authors**  
14

15  
16 Ghaeth H. Yassen, BDS, MSD, PhD,\* Alaa H.A. Sabrah, BDS, MSD, PhD, \*†  
17 George J. Eckert, MAS,‡ Jeffrey A. Platt, DDS, MS, \*  
18

19  
20 \*Division of Dental Biomaterials, Department of Restorative Dentistry, Indiana  
21 University School of Dentistry, Indianapolis, Indiana.  
22

23  
24 †Department of Conservative Dentistry, Faculty of Dentistry, The University of  
25 Jordan, Amman, Jordan.  
26

27  
28 ‡Department of Biostatistics, Indiana University School of Medicine, Indianapolis,  
29 Indiana.  
30  
31

32  
33  
34 Corresponding author:  
35

36  
37 Ghaeth H. Yassen  
38 Indiana University School of Dentistry  
39 Department of Restorative Dentistry  
40 Division of Dental Biomaterials  
41 1121 W. Michigan St.  
42 Indianapolis, IN, 46202, USA  
43  
44 Tel: +1-317-437-2240; fax: +1-317-278-7462.  
45  
46 E-mail address: gyassen@iupui.edu  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58

59 This is the author's manuscript of the article published in final edited form as:  
60 Yassen, G. H., Sabrah, A. H. A., Eckert, G. J., & Platt, J. A. (2015). Effect of Different Endodontic Regeneration  
61 Protocols on Wettability, Roughness, and Chemical Composition of Surface Dentin. *Journal of Endodontics*,  
62 41(6), 956–960. <http://doi.org/10.1016/j.joen.2015.02.023>  
63  
64  
65

- We investigated the contact angle between a blood analogue and dentin surface after various endodontic regeneration treatments.
- All endodontic regeneration protocols caused significant increase in contact angle (reduction in wettability).
- Calcium hydroxide regeneration protocol caused significant reduction in dentin wettability compared to the use of triple antibiotic paste.

## Abstract

**Introduction:** We investigated the changes in physiochemical properties of dentin surfaces after performing different endodontic regeneration protocols. **Methods:** Human dentin slices were randomized into four treatment groups and one untreated control group (n=10). One treatment group was irrigated with sodium hypochlorite (NaOCl) for five minutes, followed by ethylenediaminetetraacetic acid (EDTA) for 10 minutes. The other three treatment groups were irrigated with NaOCl, treated for four weeks with either triple antibiotic paste (TAP), diluted triple antibiotic paste (DTAP), or calcium hydroxide [Ca(OH)<sub>2</sub>], then irrigated with EDTA. After treatment, contact angles between a blood analogue and dentin surfaces were evaluated. Surface roughness and chemical composition were characterized using optical profilometry and energy dispersive X-ray spectroscopy, respectively. One-way ANOVA, followed by Fisher's Least Significant Difference tests were used for statistical analyses. **Results:** All treatment groups showed significant reduction in wettability and significant increase in surface roughness, when compared to untreated dentin. Dentin treated with Ca(OH)<sub>2</sub> had significantly lower wettability compared to all other groups. No significant difference in wettability was found between dentin treated with NaOCl+EDTA, DTAP or TAP protocols. Dentin treated with TAP had significantly higher surface roughness compared to all other groups. Untreated dentin and NaOCl+EDTA treated dentin had significantly higher calcium and phosphorus, as well as significantly lower carbon compared to dentin treated with Ca(OH)<sub>2</sub>, DTAP, and TAP. **Conclusions:** Endodontic regeneration protocols had a significant effect on wettability, surface roughness, and chemical composition of surface dentin. The Ca(OH)<sub>2</sub> protocol caused significant reduction in dentin wettability compared to TAP or DTAP protocols.

1  
2  
3  
4 **Keywords:** Endodontic regeneration, Triple antibiotic paste, Calcium hydroxide, Dentin,  
5  
6 wettability, surface roughness.  
7

## 8 **Introduction**

9  
10  
11  
12 Endodontic regeneration procedures are contemporary, biologically-based therapies that  
13  
14 manage immature teeth with necrotic pulps. These procedures may offer several advantages over  
15  
16 traditional treatments of necrotic immature teeth, such as shorter treatment time (1) and  
17  
18 continuous root development (1, 2). The first critical aspect of endodontic regeneration  
19  
20 procedures includes the disinfection of root canal systems utilizing intracanal irrigants, mainly  
21  
22 sodium hypochlorite, and medicaments (3). The most commonly used medicaments during  
23  
24 endodontic regeneration are triple antibiotic paste (TAP) and calcium hydroxide [Ca(OH)<sub>2</sub>] (3).  
25  
26 However, recent recommendations suggest the use of low concentrations of TAP, ranging from  
27  
28 0.1-1 mg/mL, to avoid cytotoxic effects against human stem cells of the apical papilla (4, 5).  
29  
30  
31  
32 Furthermore, concerns have been raised regarding the dental discoloration effect of minocyclin  
33  
34 present in TAP (6), as well as the development of antimicrobial resistance and allergic reaction  
35  
36 to antibiotic medicaments (7). The other important aspect of endodontic regeneration procedures  
37  
38 includes creating a regenerative biological environment inside the root canal system through  
39  
40 irrigation with ethylenediaminetetraacetic acid (EDTA) and the initiation of a blood clot (3, 8).  
41  
42  
43  
44  
45  
46

47 Evoking bleeding and efficiently wetting root canal dentin may improve the interaction  
48  
49 between stem cells and the dentin surface. Indeed, the induced bleeding step in regenerative  
50  
51 procedures was found to convey a significant amount of stem cells into the canal space (9). The  
52  
53 wettability of radicular dentin was suggested to modify the attachment of dental pulp cells to  
54  
55 dentin (10). Furthermore, increase in surface wettability of a substrate was associated with  
56  
57 significant improvement in cellular attachment and protein adsorption (11). The topography and  
58  
59  
60  
61  
62  
63  
64  
65

1  
2  
3  
4 chemical structure of dentin surface are surface properties that may play an important role in  
5  
6 modifying dentin wettability during endodontic regeneration. These surface properties were also  
7  
8 suggested to have significant effect on the attachment and proliferation of dental pulp stem cells  
9  
10 (12-15). This study aimed to investigate the changes in wettability, roughness, and chemical  
11  
12 structure of surface dentin after various endodontic regeneration protocols.  
13  
14

## 15 16 17 **Materials and methods**

### 18 19 20 **Sample preparation**

21  
22  
23  
24 Fifty intact human third molars were collected and stored in 0.1% thymol solution at 4 °C  
25  
26 after obtaining local IRB approval. A dentin disc (1.5 mm) was cross-sectioned from each molar  
27  
28 parallel to the occlusal surface and close to the pulp chamber, using a low-speed saw (IsoMet;  
29  
30 Buehler, Lake Bluff, IL) under constant irrigation. The non-pulpal side of each disc was flattened  
31  
32 with 500-grit silicon carbide paper (Struers, Cleveland, PA) using a polishing unit (Struers). The  
33  
34 pulpal sides of each disc was flattened using 1,200-, 2,400- and 4,000-grit silicon carbide paper  
35  
36 (Struers), followed by 0.3- $\mu$ m diamond-polishing spray (Struers). The polished specimens were  
37  
38 then sonicated in deionized water for 3 min. Deep coronal dentin, rather than radicular dentin,  
39  
40 was used to provide adequate surface area for multiple measurements of the various outcomes of  
41  
42 the study. Previous work found no significant differences between radicular and deep coronal  
43  
44 dentin in density and cross-sectional areas of dentin tubules, even after various acidic challenges  
45  
46 (16).  
47  
48  
49  
50  
51

### 52 53 **Preparation of medicaments used in the study**

54  
55  
56 A clinically recommended concentration of TAP (1000 g/mL) was prepared by mixing  
57  
58 1000 mg of United States Pharmacopeia grade antibiotic powders comprising equal portions of  
59  
60  
61

1  
2  
3  
4 metronidazole, ciprofloxacin, and minocycline (Champs Pharmacy, San Antonio, TX) with 1 mL  
5  
6 of sterile water (4, 17). A diluted paste-like consistency of 1 mg/mL TAP (DTAP) was prepared  
7  
8 as described in recent studies (18, 19). In summary, 50 mg of TAP antibiotic powders were  
9  
10 dissolved in 50 mL of sterile water. Then, 4 g of methylcellulose powder (Methocel 60 HG,  
11  
12 Sigma-Aldrich, St Louis, MO) was incorporated into the 50 mL of TAP solution under magnetic  
13  
14 stirring for 2 hours to obtain a homogenous DTAP. A commercial Ca(OH)<sub>2</sub> intracanal  
15  
16 medicament (UltraCal XS; Ultradent, South Jordan, UT) was also used in this study.  
17  
18  
19  
20

### 21 **Treatment procedure**

22  
23 The dentin discs were randomized into four treatment groups and an untreated control  
24  
25 group (n=10 per group). Samples in the control groups were stored for four weeks at 37 °C in a  
26  
27 sealed 2 mL conical sample cup (Fisher Scientific, Pittsburgh, PA, USA) at approximately 100%  
28  
29 humidity. In the first treatment group, the pulpal side of each dentin disc was slowly irrigated  
30  
31 with 20 mL of 1.5% NaOCl for five minutes using a 27-gauge needle. Samples were then stored  
32  
33 for four weeks at 37 °C in a sealed 2 mL conical sample cup at approximately 100% relative  
34  
35 humidity. After that, the pulpal side of each dentin disc was irrigated with 20 mL of 17% EDTA  
36  
37 for 10 minutes. For the other three treatment groups, the pulpal side of each dentin disc was  
38  
39 irrigated with 20 mL of 1.5% NaOCl for five minutes and treated with 0.1 mL of either TAP,  
40  
41 DTAP, or Ca(OH)<sub>2</sub> for four weeks at 37 °C in a sealed 2 mL conical sample cup at  
42  
43 approximately 100% relative humidity. After four weeks, the treated side of each dentin  
44  
45 specimen was irrigated with 20 mL of 17% EDTA for 10 minutes. The application time of  
46  
47 intracanal medicaments, as well as the irrigation time and volume of both NaOCl and EDTA,  
48  
49 were selected according to recent clinical recommendations for the endodontic regeneration  
50  
51 procedures (8).  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1  
2  
3  
4 **Blood analogue preparation and contact angle measurement**  
5

6  
7 To evaluate dentin wettability within the context of endodontic regeneration, contact  
8  
9 angle measurements between a blood analogue and dentin were performed. A solution that falls  
10  
11 within the normal range of human blood viscosity (3-4 centipoise) was prepared, as described in  
12  
13 previous studies (20, 21). In summary, 200 mL of distilled water was mixed with 100 mL of  
14  
15 100% glycerol (Sigma-Aldrich) at room temperature (22 °C) for 30 minutes under a magnetic  
16  
17 stirrer to create a blood analogue with a viscosity of 3.2 centipoise.  
18  
19

20  
21 Prior to contact angle measurements, each dentin specimen was air-dried for three  
22  
23 seconds. A Goniometer (Fibro system ab, Stockholm, Sweden) was then used to measure the  
24  
25 static contact angles between the chemically treated dentin surfaces and the blood analogue,  
26  
27 utilizing the sessile drop method. Three drops (2 µl/drop) of the blood analogue were vertically  
28  
29 dispensed on each treated dentin surface at three different locations, using a goniometer manual  
30  
31 dispensing unit (Fibro system ab). Images were captured immediately after deposition using a  
32  
33 microvideo system, and contact angles were automatically provided. All measurements were  
34  
35 performed at room temperature (22 °C). The three contact angle measurements obtained from  
36  
37 each dentin specimen were averaged to obtain a single value for each sample.  
38  
39  
40  
41

42  
43 **Surface roughness measurement.**  
44

45  
46 After contact angle measurements, each specimen was washed with 5 mL of sterile water  
47  
48 and left to dry for 10 minutes before conducting roughness analyses. Each specimen was then  
49  
50 horizontally positioned in an optical profilometer (Proscan 2000; Scantron, Venture Way,  
51  
52 Taunton, UK) and three randomly selected areas (1x1 mm<sup>2</sup>) from the treated side of each  
53  
54 specimen were scanned. The step size was set at 0.01 mm, and the number of steps were at 100  
55  
56 on both X and Y axes. Two surface roughness outcomes (Ra and Rq) were then obtained using  
57  
58  
59  
60  
61

1  
2  
3  
4 dedicated software (Proscan, 2000). Both Ra (arithmetic average roughness) and Rq (geometric  
5 average roughness obtained by calculating the root mean square roughness) were measured in  
6  
7 this study, in order to have an enhanced understanding of the dentin surface profile after various  
8  
9 treatments. The three roughness measurements obtained from each dentin specimen were  
10  
11 averaged to obtain a single value for each sample.  
12  
13

### 14 15 **Energy dispersive X-ray (EDX) measurement**

16  
17  
18  
19 After roughness measurement, five samples were randomly selected from each group for  
20  
21 EDX analyses. Each selected sample was dried for 48 h and the weight percentages of calcium  
22  
23 (Ca), phosphorus (P), carbon (C), and nitrogen (N) were measured from treated dentin surfaces  
24  
25 using scanning electron microscopy (JEOL 7800F, Peabody, MA) equipped with EDX  
26  
27 spectroscopy (EDAX Octane Super detector, Mahwah, NJ ). The samples were not sputter coated  
28  
29 to insure the precise identification of all selected elements. The EDX system was operated at 15  
30  
31 kV accelerated voltage and 1000 x magnification. EDX analyses were performed on five  
32  
33 randomly selected spots for each treated surface. The relative contribution of the four measured  
34  
35 elements was automatically normalized to a total of 100%. The five measurements of each  
36  
37 detected element from a treated dentin surface were averaged to obtain a single value.  
38  
39  
40  
41  
42

### 43 **Statistical analyses**

44  
45  
46 All data were checked for normality using the Kolmogorov-Smirnov test and a natural  
47  
48 logarithm transformation of surface roughness, phosphate, and calcium data was performed to  
49  
50 satisfy the normality assumptions. The effects of various endodontic regeneration protocols on  
51  
52 measured outcomes were examined using one-way ANOVA and Fisher's Protected Least  
53  
54 Significant Differences to control the overall significance level at 5%.  
55  
56  
57  
58  
59  
60  
61



1  
2  
3  
4 **Results**  
5

6  
7 Figure 1 shows that untreated dentin had a significantly lower contact angle than dentin  
8  
9 treated with NaOCl+EDTA (p=0.0003) as well as dentin treated with DTAP (p<.0001), TAP  
10 (p<.0001), and Ca(OH)<sub>2</sub> (p<.0001) protocols. Dentin treated with NaOCl+EDTA had a  
11 significantly lower contact angle than dentin treated with TAP (p=0.03) and Ca(OH)<sub>2</sub> (p<.0001)  
12 protocols. Furthermore, dentin treated with DTAP or TAP protocols had a significantly lower  
13 contact angle than dentin treated with Ca(OH)<sub>2</sub> (p=0.005, p=0.008).  
14  
15  
16  
17  
18  
19  
20

21  
22 Untreated dentin had significantly lower Ra and Rq than dentin treated with  
23 NaOCl+EDTA (p=0.0001) as well as dentin treated with Ca(OH)<sub>2</sub> (p=0.03 and p=0.009), DTAP  
24 (p=0.01 and p=0.008), and TAP (p<.0001) protocols (Figure2A-B). Furthermore, dentin treated  
25 with TAP had significantly higher Ra and Rq than dentin treated with NaOCl+EDTA (p=0.01  
26 and p=0.02) as well as dentin treated with Ca(OH)<sub>2</sub> (p=0.0001 and p=0.0003) or DTAP  
27 (p=0.0001 and p=0.0003).  
28  
29  
30  
31  
32  
33  
34  
35

36 Table 1 shows that dentin treated with NaOCl+EDTA had significantly higher Ca and P  
37 compared to all other groups (p<0.0001). Additionally, untreated dentin had significantly higher  
38 Ca and P compared to dentin treated with Ca(OH)<sub>2</sub> (p<0.0001), DTAP (p<0.0001), and TAP  
39 (p<0.0001). Furthermore, dentin treated with Ca(OH)<sub>2</sub> had significantly higher Ca and P  
40 compared to that treated with TAP (p<0.0001).  
41  
42  
43  
44  
45  
46  
47

48 Dentin treated with NaOCl+EDTA had significantly lower C and N (p<0.0001)  
49 compared to dentin treated with Ca(OH)<sub>2</sub>, DTAP, and TAP (Table 1). Untreated dentin had  
50 significantly lower C (p<0.0001) than dentin treated with Ca(OH)<sub>2</sub>, DTAP, and TAP. Dentin  
51 treated with Ca(OH)<sub>2</sub> and DTAP had significantly lower carbon than dentin treated with TAP  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1  
2  
3  
4 (p<0.0001). Dentin treated with TAP had significantly lower N than untreated dentin (p=0.03)  
5  
6 and dentin treated with Ca(OH)<sub>2</sub> (p=0.001) or DTAP (p=0.004).  
7  
8

## 9 **Discussion**

10  
11 The presence of an intimate contact between stimulated bleeding and root canal dentin  
12  
13 during endodontic regeneration may improve the formation of a blood clot-dentin natural  
14  
15 scaffold, accelerate the interaction between dentin growth factors and blood containing stem  
16  
17 cells, and improve the deposition of new dentin. Wettability can be expressed in terms of contact  
18  
19 angle, which measures the ability of a liquid to spread on a plane solid surface. The contact angle  
20  
21 has an inverse relationship with wettability. Furthermore, dentin wettability is greatly dependent  
22  
23 on chemical structure and surface roughness of the surface (22, 23). Therefore, the wettability,  
24  
25 topography and chemical composition of surface dentin were investigated in the current study.  
26  
27  
28  
29  
30

31  
32 Our results demonstrated a significant decrease in dentin wettability among all treatment  
33  
34 groups when compared with untreated control dentin. This could be explained by the significant  
35  
36 increase in surface roughness outcomes reported among all treatment groups. Recent studies  
37  
38 suggested that lower dentin wettability was associated with higher surface roughness (22, 24).  
39  
40 The significantly lower dentin wettability after TAP, DTAP and Ca(OH)<sub>2</sub> treatment can also be  
41  
42 explained by the significant reduction in the inorganic phase represented by Ca and P and the  
43  
44 significant increase in the organic phase represented by C among these groups. The acidic nature  
45  
46 of TAP (pH=2.9) (25) and EDTA's calcium chelating ability may be responsible for the net  
47  
48 surface demineralization effect of dentin surfaces after various regeneration protocols. It is well  
49  
50 established that the inorganic hydroxyapatite phase of dentin has high wettability while the  
51  
52 organic collagen phase of dentin has low wettability (26). Indeed, a previous study reported a  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1  
2  
3  
4 correlation between increases in dentin wettability and higher concentrations of calcium on the  
5  
6 dentin surface (27).  
7  
8  
9

10 An interesting finding of this study is that dentin treated with  $\text{Ca(OH)}_2$  showed  
11 significantly lower wettability compared to all other groups. This relatively poor adherence may  
12 support the finding of a recent clinical study that found that endodontic regeneration cases  
13 disinfected with  $\text{Ca(OH)}_2$  had significantly thinner root walls compared to regeneration cases  
14 disinfected with TAP (2). The results of dentin surface topography and chemical composition in  
15 our study may not be enough to completely justify the reported low dentin wettability after  
16  $\text{Ca(OH)}_2$  treatment. One additional explanation is the effect of  $\text{Ca(OH)}_2$  on surface dentin  
17 modulus of elasticity. A recent study proposed that dentin wettability was affected by changes in  
18 surface dentin modulus of elasticity (23) and  $\text{Ca(OH)}_2$  was found to significantly change dentin  
19 modulus of elasticity after one week of application (28).  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34

35 In the current study, a dentin surface treated with NaOCl+EDTA showed significantly  
36 higher inorganic phase (Ca and P) and significantly lower organic phase (C and N) compared to  
37 untreated control dentin. This agrees with previous work that reported significantly higher  
38 inorganic composition of dentin treated with EDTA and NaOCl compared to untreated dentin  
39 (29). This could be explained by the presence of smear layer on untreated control dentin that is  
40 rich with organic debris represented by N and C (30). On the other hand, the final 10 minutes of  
41 irrigation with EDTA in dentin-treated NaOCl+EDTA is expected to remove the smear layer. A  
42 relatively recent study suggested that root canal irrigation with NaOCl followed by EDTA  
43 caused complete removal of the smear layer (31). Despite the high inorganic phase in surface  
44 dentin treated with NaOCl+EDTA, a significantly lower wettability of dentin irrigated with  
45 NaOCl+EDTA compared to untreated control dentin was observed in this study. This agrees with  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1  
2  
3  
4 a previous study that suggested significantly lower wettability of dentin irrigated with EDTA  
5  
6 compared to untreated dentin (32). This can be explained by the ability of EDTA to increase  
7  
8 surface roughness, completely remove the smear layer and create wide open dentinal tubules (33,  
9  
10 34). Open dentin tubules with larger diameters have been proposed to significantly decrease the  
11  
12 dentin wettability due to the effect of small air pockets inside the dentin tubules (22-24).  
13  
14  
15  
16

17 A Newtonian blood surrogate was used in the current study instead of non-Newtonian  
18  
19 human blood to detect the wettability of dentin. This standardized approach was used in an  
20  
21 attempt to explore the influence of change in physiochemical surface properties of dentin after  
22  
23 various regeneration protocols on the static contact angle. The measurement of dynamic contact  
24  
25 angles using actual human blood in future studies may provide further understanding of the  
26  
27 spreading behavior of blood on dentin surfaces.  
28  
29  
30  
31  
32

33 Collectively, our study showed that the endodontic regeneration protocols had a  
34  
35 significant effect on dentin surface wettability, roughness, and chemical composition. The use of  
36  
37 Ca(OH)<sub>2</sub> during endodontic regeneration caused a significant reduction in dentin wettability  
38  
39 compared to the use of TAP or DTAP intracanal medicaments. Furthermore, TAP caused  
40  
41 significant increase in surface roughness compared to other tested intracanal medicaments.  
42  
43  
44

#### 45 **Acknowledgments**

46  
47  
48 The authors deny any conflicts of interest related to this study.  
49

#### 50 **References**

- 51  
52  
53  
54 1. Jeeruphan T, Jantarat J, Yanpiset K, et al. Mahidol study 1: comparison of radiographic  
55 and survival outcomes of immature teeth treated with either regenerative endodontic or  
56 apexification methods: a retrospective study. J Endod 2012;38:1330-6.  
57  
58 2. Bose R, Nummikoski P, Hargreaves K. A retrospective evaluation of radiographic  
59 outcomes in immature teeth with necrotic root canal systems treated with regenerative  
60 endodontic procedures. J Endod 2009;35:1343-9.  
61

3. Diogenes A, Henry MA, Teixeira FB, et al. An update on clinical regenerative endodontics. *Endod Top* 2013;28:2-23.
4. Ruparel NB, Teixeira FB, Ferraz CC, et al. Direct effect of intracanal medicaments on survival of stem cells of the apical papilla. *J Endod* 2012;38:1372-5.
5. Althumairy RI, Teixeira FB, Diogenes A. Effect of dentin conditioning with intracanal medicaments on survival of stem cells of apical papilla. *J Endod* 2014;40:521-5.
6. Nagata JY, Gomes BP, Rocha Lima TF, et al. Traumatized immature teeth treated with 2 protocols of pulp revascularization. *J Endod* 2014;40:606-12.
7. Reynolds K, Johnson JD, Cohenca N. Pulp revascularization of necrotic bilateral bicuspid using a modified novel technique to eliminate potential coronal discoloration: a case report. *Int Endod J* 2009;42:84-92.
8. Diogenes AR, Ruparel NB, Teixeira FB, et al. Translational science in disinfection for regenerative endodontics. *J Endod* 2014;40:S52-7.
9. Lovelace TW, Henry MA, Hargreaves KM, et al. Evaluation of the delivery of mesenchymal stem cells into the root canal space of necrotic immature teeth after clinical regenerative endodontic procedure. *J Endod* 2011;37:133-8.
10. Huang X, Zhang J, Huang C, et al. Effect of intracanal dentine wettability on human dental pulp cell attachment. *Int Endod J* 2012;45:346-53.
11. Wei J, Igarashi T, Okumori N, et al. Influence of surface wettability on competitive protein adsorption and initial attachment of osteoblasts. *Biomed Mater* 2009;4:045002.
12. Liu J, Jin TC, Chang S, et al. Adhesion and growth of dental pulp stem cells on enamel-like fluorapatite surfaces. *J Biomed Mater Res A* 2011;96:528-34.
13. Carvalho A, Pelaez-Vargas A, Gallego-Perez D, et al. Micropatterned silica thin films with nanohydroxyapatite micro-aggregates for guided tissue regeneration. *Dent Mater* 2012;28:1250-60.
14. Collart-Dutilleul PY, Secret E, Panayotov I, et al. Adhesion and proliferation of human mesenchymal stem cells from dental pulp on porous silicon scaffolds. *ACS Appl Mater Interfaces* 2014;6:1719-28.
15. Kolind K, Kraft D, Boggild T, et al. Control of proliferation and osteogenic differentiation of human dental-pulp-derived stem cells by distinct surface structures. *Acta Biomater* 2014;10:641-50.
16. Caiado AC, de Goes MF, de Souza-Filho FJ, et al. The effect of acid etchant type and dentin location on tubular density and dimension. *J Prosthet Dent* 2010;103:352-61.
17. Berkhoff JA, Chen PB, Teixeira FB, et al. Evaluation of triple antibiotic paste removal by different irrigation procedures. *J Endod* 2014;40:1172-7.
18. Prather BT, Ehrlich Y, Spolnik K, et al. Effects of two combinations of triple antibiotic paste used in endodontic regeneration on root microhardness and chemical structure of radicular dentine. *J Oral Sci* 2014;56:245-51.
19. Yassen GH, Eckert GJ, Platt JA. Effect of intracanal medicaments used in endodontic regeneration procedures on microhardness and chemical structure of dentin. *Restor Dent Endod* 2015 (In Press)
20. McGillicuddy JW, Chambers SD, Galligan DT, et al. In vitro fluid mechanical effects of thoracic artificial lung compliance. *ASAIO J* 2005;51:789-94.
21. Schewe RE, Khanfer KM, Arab A, et al. Design and in vitro assessment of an improved, low-resistance compliant thoracic artificial lung. *ASAIO J* 2012;58:583-9.

22. Ramos SM, Alderete L, Farge P. Dentinal tubules driven wetting of dentin: Cassie-Baxter modelling. *Eur Phys J E Soft Matter* 2009;30:187-95.
23. Pelin IM, Trunfio-Sfarghiu AM, Farge P, et al. Multiscale characterization of partially demineralized superficial and deep dentin surfaces. *Eur J Oral Sci* 2013;121:341-8.
24. Farge P, Alderete L, Ramos SM. Dentin wetting by three adhesive systems: influence of etching time, temperature and relative humidity. *J Dent* 2010;38:698-706.
25. Yassen GH, Chu TM, Eckert G, et al. Effect of medicaments used in endodontic regeneration technique on the chemical structure of human immature radicular dentin: an in vitro study. *J Endod* 2013;39:269-73.
26. Eliades G. Clinical relevance of the formulation and testing of dentine bonding systems. *J Dent* 1994;22:73-81.
27. Panighi M, G'Sell C. Influence of calcium concentration on the dentin wettability by an adhesive. *J Biomed Mater Res* 1992;26:1081-9.
28. Kawamoto R, Kurokawa H, Takubo C, et al. Change in elastic modulus of bovine dentine with exposure to a calcium hydroxide paste. *J Dent* 2008;36:959-64.
29. Dogan H, Qalt S. Effects of chelating agents and sodium hypochlorite on mineral content of root dentin. *J Endod* 2001;27:578-80.
30. Eick JD, Wilko RA, Anderson CH, et al. Scanning electron microscopy of cut tooth surfaces and identification of debris by use of the electron microprobe. *J Dent Res* 1970;49:Suppl:1359-68.
31. Lotfi M, Vosoughhosseini S, Saghiri MA, et al. Effect of MTAD as a final rinse on removal of smear layer in ten-minute preparation time. *J Endod* 2012;38:1391-4.
32. Dogan Buzoglu H, Calt S, Gumusderelioglu M. Evaluation of the surface free energy on root canal dentine walls treated with chelating agents and NaOCl. *Int Endod J* 2007;40:18-24.
33. Calt S, Serper A. Time-dependent effects of EDTA on dentin structures. *J Endod* 2002;28:17-9.
34. Teixeira CS, Felipe MC, Felipe WT. The effect of application time of EDTA and NaOCl on intracanal smear layer removal: an SEM analysis. *Int Endod J* 2005;38:285-90.

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

Figure 1. Mean (SE) of contact angles measured on dentin surfaces after various endodontic regeneration protocols.

Figure 2A. Mean (SE) of Ra roughness outcome measured on dentin surfaces after various endodontic regeneration protocols.

Figure 2B. Mean (SE) of Rq roughness outcome measured on dentin surfaces after various endodontic regeneration protocols.

Figure 1  
[Click here to download high resolution image](#)

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49

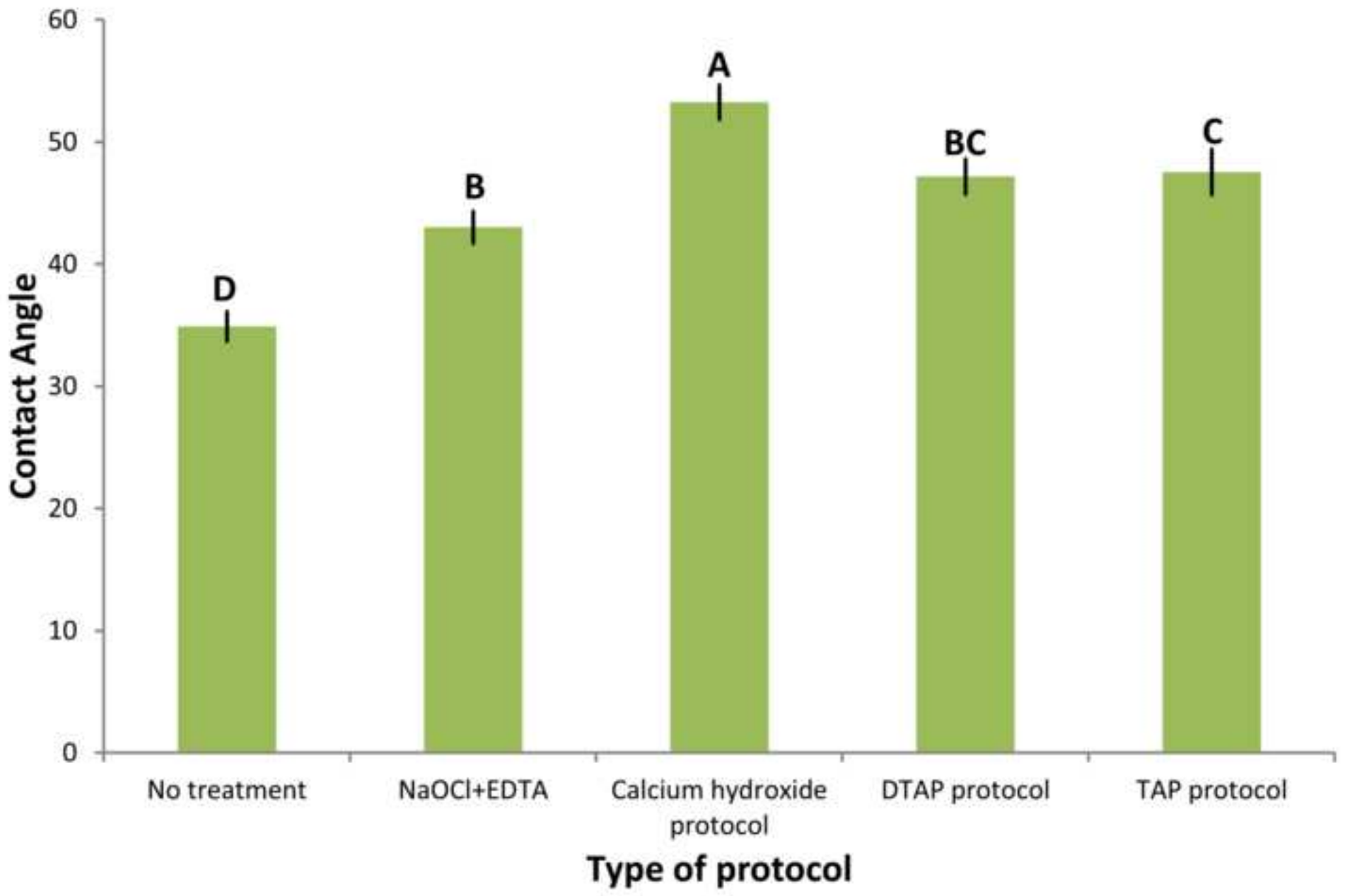
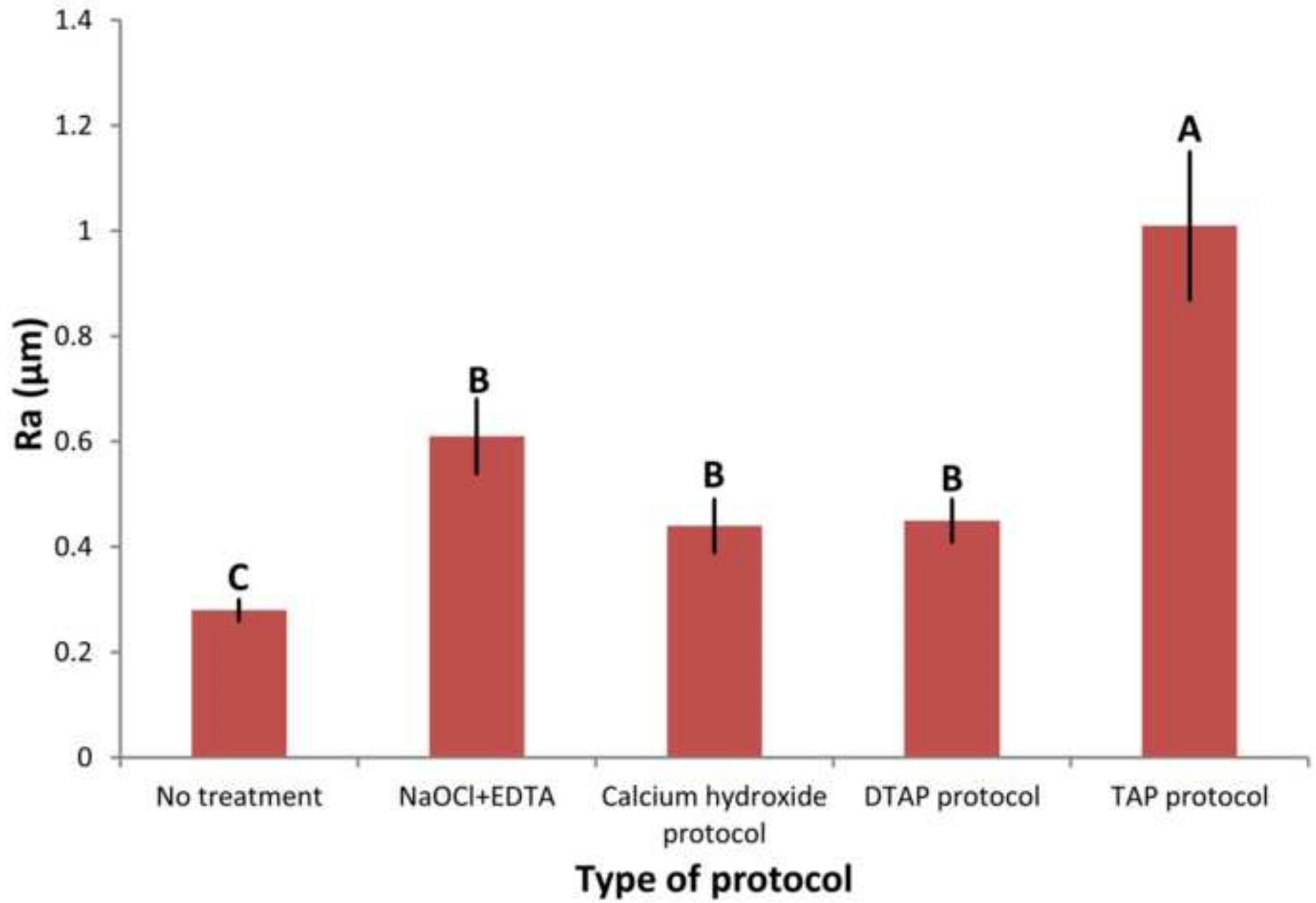




Figure 2A  
[Click here to download high resolution image](#)



1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49

**Figure 2B**  
[Click here to download high resolution image](#)

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49

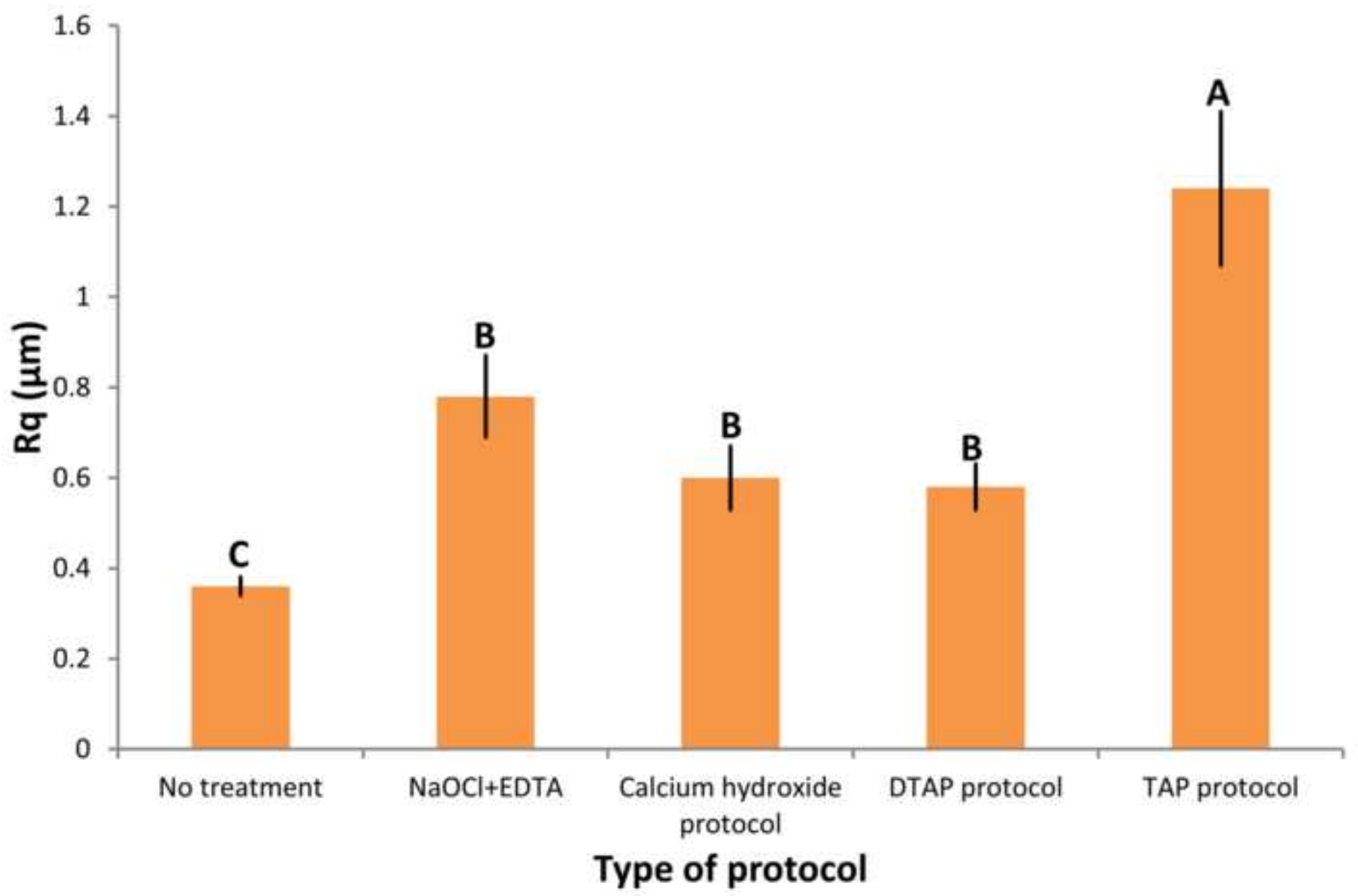


Table 1. Mean (SE) of weight percentages of chemical elements measured on dentin surfaces after various endodontic regeneration protocols.

Type of treatment*	Calcium	Phosphorus	Nitrogen	Carbon
NaOCl + EDTA	46.33 (3.8)A	22.1 (0.8)A	19 (3)C	12 (1.8)D
Untreated dentin	11.4 (2.2)B	8.7 (1)B	57(2.4)A	23 (0.8)C
Ca(OH) <sub>2</sub> Protocol	0.3 (0.04)C	0.6 (0.02)C	61(1.9)A	38 (1.1)B
DTAP Protocol	0.2(0.01)D	0.51(0.03)C	60(0.7)A	40 (1.3)B
TAP Protocol	0.04 (0.01)E	0.2 (0.02)D	50(1.2)B	49 (1.1)A

\*Within each element, different upper-case letter indicate a significant difference between various endodontic regeneration protocols